

Lyo NZYSupreme One-step RT-qPCR Probe Master Mix (2x)

Catalogue number:

MB41501, for 1,5 ml (150 x 20 µL)

Description

Lyo NZYSupreme One-step RT-qPCR Probe Master Mix is a freeze-dried reaction mixture developed for first-strand cDNA synthesis and subsequent real-time PCR in a single tube. This offers great convenience and minimize the risk of errors and contaminations. The master mix, formulated as a 2x reaction mixture, is provided with an optimized reconstitution buffer designed to confer maximal levels of stability and enzymatic activity. After reconstitution, the mixture contains all components necessary for both cDNA synthesis and real-time PCR (including enzymes, dNTPs, stabilizers and enhancers), except primers, probes and RNA template. This master mix was engineered with a dual hot-start enzyme control mechanism to provide the highest detection sensitivity. In addition, the latest developments in PCR enhancers have been incorporated in the Lyo NZYSupreme One-step RT-qPCR Probe Master Mix, including buffer chemistry and incorporation of highly robust engineered enzymes. This master mix does not contain ROX and it was specifically developed for probe-detection technology. For qPCR instruments that require ROX reference dye, please add ROX (Cat. No. MB406) according to the table presented in the section "ROX reference dye".

Storage temperature

Lyo NZYSupreme One-step RT-qPCR Probe Master Mix can be shipped in a range of temperatures from dry ice to room-temperature. Although the product components provided are stable at room temperature, they should be stored at -20 °C upon arrival. Short-term storage for 1-2 weeks at room temperature will not compromise the performance of the product. Once the lyophilized master mix has been resuspended, it should be stored -20 °C in a constant temperature freezer. Minimize the number of freeze-thaw cycles by storing in working aliquots. The master mix will remain stable till the expiry date if stored as specified.

Pack components

Component	MB41501
Lyo NZYSupreme One-step RT-qPCR Probe Master Mix (2x)	3 tubes with lyophilized master mix
RT-qPCR master mix reconstitution buffer	1.75 mL

Preparation of Lyo NZYSupreme One-step RT-qPCR Probe Master Mix

Reconstitute each tube of Lyo NZYSupreme One-step RT-qPCR Probe Master Mix with 525 µL of the RT-qPCR master mix reconstitution buffer provided. Complete hydration of the mixture might take 2-3 min. Mix gently by flicking the tube, and spin down. Do not replace the reconstitution buffer with water or any other

buffer. The master mix is then ready to use as a 2x One-step RT-qPCR master mix.

Compatible real-time PCR instruments

The master mix is compatible with real-time PCR instruments that do not require a passive reference signal for data normalization. It has been optimized to be compatible with the following real-time PCR instruments:

Bio-Rad®:

CFX96™; CFX384™; iCycler®; iQ™5; Opticon™; Opticon™ 2

Qiagen (Corbett):

Rotor-Gene™ 3000; Rotor-Gene™ 6000 & Rotor-Gene™ Q

Roche:

Lightcycler® 96; Lightcycler® 480 & Lightcycler® Nano

Applied Biosystems (with optional ROX addition):

7000; 7300; 7700; 7900; 7900HT; 7900HT FAST; StepOne™ & StepOne™plus; 7500; 7500 FAST; QuantStudio™ 6, 7, 12k Flex & ViiA7™

Protocol

The following protocol serves as a general guideline and a starting point for any One-step RT-qPCR procedure. Optimal reaction conditions (e.g. incubation times, temperatures and template concentration) may vary and, in particular conditions, may require further optimization.

RT-qPCR reaction set-up: the given volumes are based on a standard 20 µL final reaction mix and can be scale adjusted.

Lyo NZYSupreme One-step RT-qPCR Probe Master Mix (2x) ^(*)	10 µL	1x
10 µM forward primer	0.8 µL	400 nM ^(*)
10 µM reverse primer	0.8 µL	400 nM ^(*)
10 µM probe	0.2 µL	100 nM ^(*)
Template	up to 8.2 µL	-
Nuclease-free water	up to 20 µL	-

^(*) The master mix does not contain ROX, but, if required, addition of this internal passive reference dye can be conducted in a separate step. The final concentration will vary according to the qPCR instrument used. Please follow instructions described in the section "ROX reference dye" below.

^(*) See section of "General considerations" below for more details about primers and probes final concentrations in the reaction.

Testing and Ct values: When comparing this RT-qPCR master mix with a mix from another supplier we strongly recommend amplifying from a 10-fold template dilution series. Loss of detection at low template concentration is the only direct measurement of sensitivity. An early Ct value is not an indication of good sensitivity, but rather an indication of speed.

ROX reference dye: Lyo NZYSupreme One-step RT-qPCR Probe Master Mix (2x) is compatible with the majority of thermocyclers available in the market and can include ROX passive reference dye to normalize non-PCR-related fluctuations in fluorescence. If ROX addition is required for your qPCR platform, an optimal quantity of this dye should be included in your master mix. ROX can be purchased separately from NZYTech (Cat. No. MB406) in a ready-to-use solution of 25 µM. The recommended amount of ROX for the most common qPCR instruments is stated in the table below:

qPCR instrument	Volume of ROX per 525 µL of 2x master mix (2x) (*)
Applied Biosystems: 7000/7300/7700/7900/7900HT/7900HT FAST/StepOne™/ StepOne™plus	15 µL
Applied Biosystems: 7500/7500FAST/QuantStudio™ 6, 7, 12k Flex/ViiA7™	2 µL
Bio-Rad®: CFX96™/CFX384™/iCycler®/iQ™5/Opticon™/ Opticon™ 2 Qiagen: Rotor-Gene™ 3000/6000/Q Roche: Lightcycler® 96/480/Nano	Not required

(*) For different volumes please scale-up or scale-down the volume of ROX accordingly.

Suggested thermal cycling conditions

Lyo NZYSupreme One-step RT-qPCR Probe Master Mix was optimized for the amplification of RNA fragments up to 200 bp under different RT-qPCR cycling conditions. The table below displays a standard 2-step cycling setup optimized on a number of platforms. However, these conditions may be adapted to suit different machine-specific protocols.

Cycles	Temp.	Time	Main reaction
1	50 °C	20 min	Reverse Transcription
1	95 °C	2-5 min	Polymerase activation
40	95 °C	5 s	Denaturation
	60 °C	30 s-50 s	Annealing/Extension

General considerations

The inherent chemical instability of RNA and the ubiquitous presence of RNases require that particular care should be taken while working with RNA. We recommend using RNase-free plasticware/reagents and work in an RNase-free area (RNase Cleaner, Cat. No. MB16001, can help removing RNases from surfaces and materials). In addition, to help prevent any carry-over DNA contamination, you should assign independent areas for reaction set-up, PCR amplification and any post-PCR gel analysis. It is essential that any tubes containing amplified PCR product are not opened in the PCR set-up area.

Primers and probe: These guidelines refer to the design and set-up of dual labeled probes. Please refer to the relevant literature when using other probe types. The specific amplification, yield and overall efficiency of any real-time RT-PCR can be critically affected by the sequence and concentration of the probes and primers, as well as by the amplicon length. We strongly recommend taking the following points into consideration when designing and running your real-time RT-qPCR experiment:

- Primers should have a melting temperature (T_m) of approximately 60 °C. The probe T_m should be approximately 10°C higher than that of the primers;
- The fragment to amplify should be between 80-200 bp in length and not superior to 300 bp;
- Final primer concentrations of 400 nM are suitable for most probe-based reactions. However, to determine the optimal concentration we recommend titrating in the range 0.2-1 µM. Forward and reverse primers concentration should be equimolar;
- A final probe concentration of 100 nM is suitable for most applications; we recommend that the final probe concentration

is at least two-fold lower than the primer concentration; to determine the optimal concentration we recommend titrating in the range 0.1-0.25 µM;

- For multiplex RT-qPCR, probe concentrations in excess of 100 nM can result in cross channel fluorescence.

Template: It is important that the RNA template is purified and devoid of contamination by RT-qPCR inhibitors (e.g. EDTA). The recommended amount of template is dependent upon the type of RNA used. Please consider the following points when selecting RNA templates:

- **Total RNA:** purified total RNA can be used in the range from 1 pg to 1 µg per 20 µL reaction;
- **mRNA:** purified mRNA can be used from 0.01 pg per 20 µL reaction.

To obtain high yield of highly purified RNA we suggest using the NZY Total RNA Isolation Kit (Cat. No. MB134).

MgCl₂: It is not necessary to supplement the reaction mixture with MgCl₂ as the Lyo NZYSupreme One-step RT-qPCR Probe Master Mix already contains an optimized concentration of MgCl₂.

RT-qPCR optimization: It may be necessary to improve the efficiency of some reactions, such as multiplexing with more than two probes, or if the target amplicon is longer than 200 bp. In these cases, the reverse transcription reaction time can be extended up to 30 minutes; the annealing/extension time can be extended up to 60 seconds.

Quality control assays

Genomic DNA contamination

Lyo NZYSupreme One-step RT-qPCR Probe Master Mix must be free of any detectable genomic DNA contamination as evaluated through a real-time PCR assay.

Nuclease assays

To test for DNase contamination, 0.2-0.3 µg of pNZY28 plasmid DNA are incubated with the master mix for 14-16 h at 37 °C. To test for RNase contamination, 1 µg of RNA is incubated with the master mix for 1 h at 37 °C. Following incubation, the nucleic acids are visualized on a GreenSafe-stained agarose gel. There must be no visible nicking or cutting of the nucleic acids.

Functional assay

Lyo NZYSupreme One-step RT-qPCR Probe Master Mix is extensively tested for activity, processivity, efficiency, sensitivity and heat activation.

V2001

Certificate of Analysis	
Test	Result
Genomic DNA contamination	Pass
Nuclease assays	Pass
Functional assay	Pass
Approved by:  Patrícia Ponte Senior Manager, Quality Systems	

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